

Digeneans Parasitic in Freshwater Fishes (Osteichthyes) of Japan. I. Aporocotylidae, Bivesiculidae and Haploporidae

Takeshi Shimazu

10486–2 Hotaka-Ariake, Azumino, Nagano 399–8301, Japan
E-mail: azygia79@gmail.com

(Received 8 August 2013; accepted 16 October 2013)

Abstract Adult digenetic trematodes (digeneans) parasitic in freshwater fishes (Osteichthyes) of Japan are reviewed in a series of several papers. The current knowledge of each of the digenean species is provided from the existing specimens and literature, including a morphological description with figures, lists of its synonyms, hosts, localities and existing specimens, and a summary of its life cycle where known. The Aporocotylidae Odhner, 1912 (*Sanguinicola uguī*, *Sanguinicola hasegawai* sp. nov. and *Sanguinicola* sp.), Bivesiculidae Yamaguti, 1934 (*Bivesicula* sp.) and Haploporidae Nicoll, 1914 (*Carassotrema koreanum*) are reviewed. *Sanguinicola hasegawai* sp. nov. is described and figured on the basis of specimens found in the blood vessels and heart of *Barbatula toni* (Dybowski, 1869) (Nemacheilidae) from Hokkaido.

Key words: digeneans, freshwater fishes, Japan, review, Aporocotylidae, Bivesiculidae, Haploporidae, *Sanguinicola hasegawai* sp. nov.

Introduction

I wrote two reviews on adult digenetic trematodes (digeneans) parasitic in freshwater fishes (Osteichthyes) of Japan (Shimazu, 1999, 2003). Since then, several papers of taxonomic importance have been published in this field. The two previous reviews contain neither morphological descriptions nor figures for any of the digenean species.

This new review will be published in a series of several papers. The current knowledge of each of the digenean species from the existing specimens and literature will be provided, including a morphological description with figures, lists of its synonyms, hosts, localities and existing specimens and a summary of its life cycle where known. Keys to genera in families and species in genera will be given if necessary. In a final paper, a general discussion on the digenean fauna of Japanese freshwater fishes, host-parasite and parasite-host lists and a key to the families will be added.

This is a first paper in the series, dealing with the Aporocotylidae Odhner, 1912, formerly the Sanguinicolidae von Graff, 1907 *sensu* Smith (2002b) in the Schistosomatoidea Stiles and Hassall, 1898 *sensu* Smith (2002a); the Bivesiculidae Yamaguti, 1934 in the Bivesiculoidea Yamaguti, 1934 *sensu* Cribb (2002); and the Haploporidae Nicoll, 1914 *sensu* Overstreet and Curran (2005) in the Haploporoidea Nicoll, 1914 *sensu* Jones (2005). The literature search ended in August 2013.

In Japan, 150–170 species of freshwater fishes (mostly Osteichthyes) are distributed (Goto, 1987). *Exorchis oviformis* Kobayashi, 1915 was the first adult digenean that was reported from a freshwater fish in Japan (Shimazu, 1999, 2003). Since then, adult digeneans of Japanese freshwater fishes have been studied by many workers (e.g., H. Kobayashi, T. Fujita, Y. Ozaki, S. Yamaguti, S. Takahashi, S. Eguchi, T. Hasegawa, N. Ishii, T. Fujino, F. Moravec, T. Shimazu, M. Urabe and others). Their published papers will be cited in the respective species of digeneans (see also

Shimazu, 1999, 2003). Early studies, which were devoted chiefly on taxonomy and life cycle, prospered after 1915, reached to the heyday in early 1920 to late 1930, and have been gradually declining since (Shimazu, 1999, 2003). Y. Ozaki's works were compiled by Shimazu (1995); and S. Yamaguti's, by Kamegai and Ichihara (1972). Shimazu (1999, 2003) counted at least 41 species in 20 genera and 12 families of digeneans reported until then. Shimazu *et al.* (2011) recorded 24 previously known, 2 new and 4 unidentified species in 17 genera and 12 families of digeneans from 42 species and 5 unidentified species of fishes from the Lake Biwa basin, Shiga Prefecture. Studies of the taxonomy, phylogenetic relationship, life cycle, geographical distribution and others of some species of digeneans have recently been in progress from not only morphological but also molecular data. In addition, cercariae and metacercariae have been extensively studied in Japan from the viewpoint of medical importance (Ito, 1964; Komiya, 1965). Further studies of them would prove that the names of some of them have preoccupied the names of some adult digeneans, e.g., *Cercaria gotoi* Ariake, 1922 and *Azygia anguillae* Ozaki, 1924 in *Azygia gotoi* (Ariake, 1922) Shimazu, 1979. In *Life cycle* of each species, the life cycle will be summarized where known, and a possible cercaria or metacercaria will be referred to.

During the above-mentioned studies, many adult digeneans were described as new species. Holotypes were designated for some of them, but not for others. Many of the original specimens including syntypes and holotypes exist: Dr. T. Fujita's Collection (Fujita's Collection) deposited in the Hokkaido University Natural History Museum (HUNHM), Sapporo, Hokkaido; Dr. S. Yamaguti's Collection (Yamaguti's Collection) and Dr. Y. Ozaki's Collection (Ozaki's Collection) in the Meguro Parasitological Museum (MPM), Tokyo; and the collections of T. Shimazu and M. Urabe in the National Museum of Nature and Science (NMNS, collection code name NSMT-PI), Tsukuba, Ibaraki Prefecture, formerly the National Science Museum, Tokyo.

Y. Ozaki deposited a total of 38 holotypes of his new species of platyhelminth parasites including digeneans in the collection of the Zoological Institute, Science Faculty, Tokyo Imperial University, Tokyo (Shimazu, 1995). This collection has been transferred to the Department of Zoology, The University Museum, The University of Tokyo, Tokyo. The latter collection today includes no holotypes of helminth parasite species including Ozaki's ones (Shimazu and Araki, 2006). Obviously, all the holotypes of helminth parasite species deposited in the Zoological Institute were lost. Furthermore, none of the original specimens of some other workers could be traced for reexamination. Probably, they were also lost (see also Shimazu *et al.*, 2011). Neotypes will be designated for some digeneans species if necessary and possible in this work.

Materials and Methods

In the present work, the classification of the subclass Digenea Carus, 1863 in the class Trematoda Rudolphi, 1808 is fundamentally based on that in the series entitled "Keys to the Trematoda" (Volume 1 edited by Gibson *et al.*, 2002; Volume 2 edited by Jones *et al.*, 2005; and Volume 3 edited by Bray *et al.*, 2008). Some of the technical terms used are different from those used in the series. Most of the family and species names of host fishes are given according to Nakabo (2013) and Eschmeyer (2013).

Synonym lists contain only the references for the original descriptions, subsequent descriptions of taxonomic importance from Japan in particular and establishments of new combinations. In *Geographical distribution*, only records of immature (juvenile) and mature (adult) worms from freshwater fishes are presented. The locality name "Japan" means that parasites were certainly obtained in Japan, but their exact localities have not yet been determined.

Specimens had been prepared by various methods, either whole-mounted in Canada balsam or serially sectioned. Many of them were borrowed from Fujita's Collection (HUNHM),

Yamaguti's and Ozaki's Collections (MPM), the collection of the NMNS, Tokyo then, and the personal collection of M. Urabe. I also used some of the specimens deposited in the Lake Biwa Museum (LBM), Kusatsu, Shiga Prefecture, for this review while preparing the paper by Shimazu *et al.* (2011). Newly collected specimens of mine were also used, and they have been deposited in the NMNS.

Measurements (length by width) are given in millimeters unless otherwise stated. The body size is indicated as follows: very small, -1 mm long; small, 1–3; fairly small, 3–7; medium, 7–12; fairly large, 12–20; large, 20–35; and very large, 35– (Dawes, 1956). The sucker width ratio is the ratio of the width of the oral sucker to that of the ventral sucker. Most of the drawings were made with the aid of a camera lucida.

The name of the Japanese journal "Dobutsugaku Zasshi" is cited as merely "Dobutsugaku Zasshi" in this work, though it was cited as "Zoological Magazine (Japan) (Dobutsugaku Zasshi)" in some of my previous papers.

Class **Trematoda** Rudolphi, 1808

Subclass **Digenea** Carus, 1863

Superfamily **Schistosomatoidea** Stiles and Hassall, 1898

Family **Aporocotylidae** Odhner, 1912

Genus **Sanguinicola** Plehn, 1905

Sanguinicola ugui Shimazu, 2007

(Figs. 1–4)

Sanguinicola ugui Shimazu, 2007: 2–4, figs. 1–6.

Host in Japan. *Tribolodon hakonensis* (Günther, 1877) (Cyprinidae) (type host) (Shimazu, 2007; this paper).

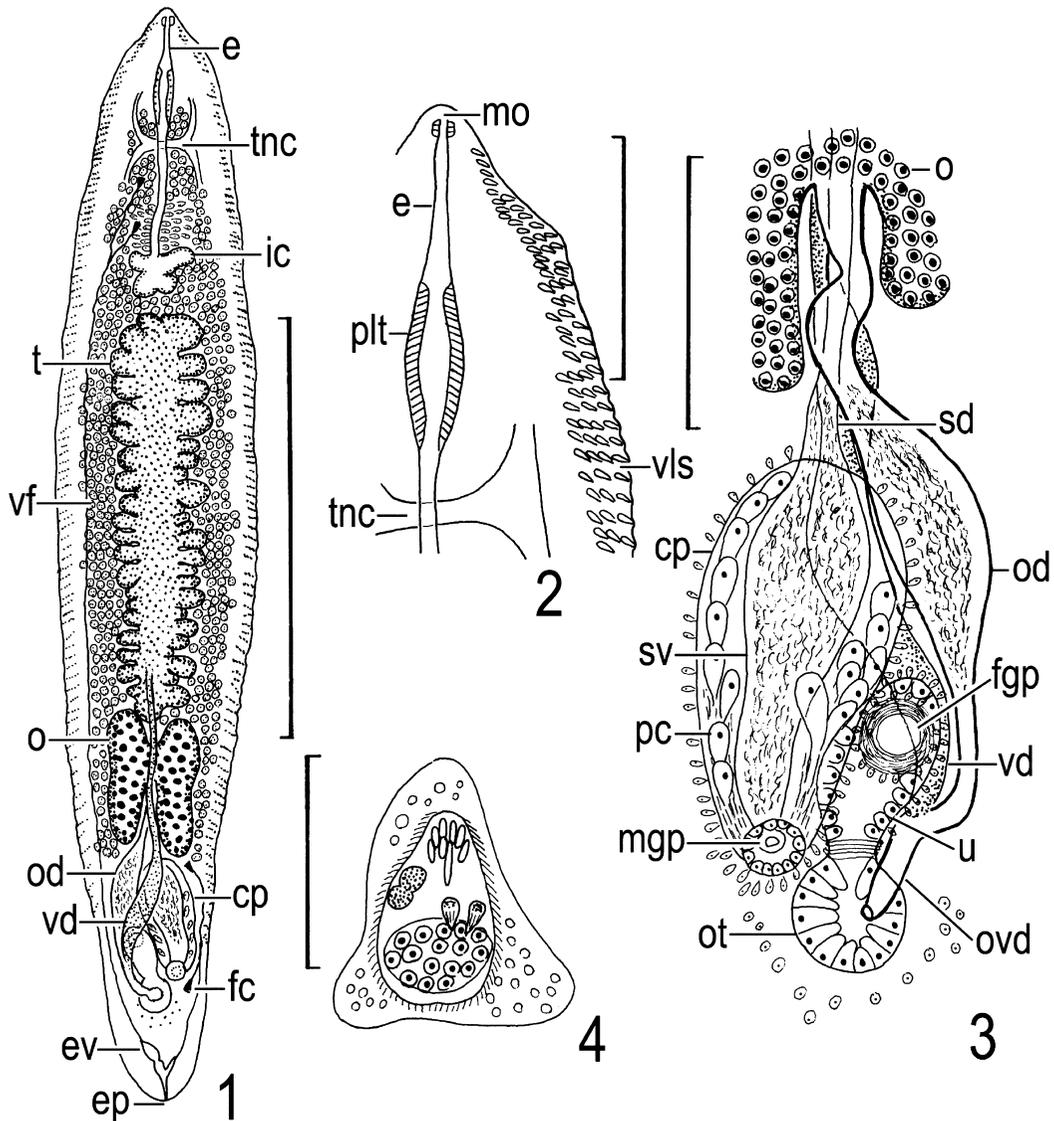
Sites of infection. Blood vessels and heart: afferent branchial artery of gills, thick branches of hepatic artery of liver, and undetermined blood vessel of kidneys; and arterial bulb of heart (Shimazu, 2007; this paper).

Geographical distribution. Nagano Prefecture: Hiroi River (type locality) at Kotobuki,

Iiyama City; Lake Suwa at Suwa City; Tenryu River at Ina City; and Sai River at Akashina-Nakagawate, Azumino City (Shimazu, 2007; this paper).

Material examined. (1) Type series of *Sanguinicola ugui*: holotype (NSMT-PI 5284) and 27 paratypes (NSMT-PI 5284–5286, 5288), adult, whole-mounted, ex afferent [not efferent] branchial artery of *Tribolodon hakonensis*, Hiroi River, 20 July 1996, 23 October 1996, 24 November 1996, 5 November 2004 (Shimazu, 2007). (2) Many specimens (NSMT-PI 5283–5295) of *S. ugui*, adult, whole-mounted, ex afferent [not efferent] branchial artery and arterial bulb [not ventricle] of *T. hakonensis*, Hiroi River, 1996–2004; 1 in infected afferent [not efferent] branchial artery (NSMT-PI 5283), adult; and 1 (NSMT-PI 5291), adult, ex arterial bulb [not ventricle] (Shimazu, 2007). (3) Some (NSMT-PI 5296–5297) of *S. ugui*, adult, ex afferent [not efferent] branchial artery of *T. hakonensis*, Lake Suwa, 2 August 1996, 5 October 1996; and some (NSMT-PI 5298) of *S. ugui*, adult, ex afferent [not efferent] branchial artery of *T. hakonensis*, Tenryu River, 9 September 2000 (Shimazu, 2007). (4) 3 (NSMT-PI 5891) and 9 (NSMT-PI 5892), adult, whole-mounted, ex afferent branchial artery, *T. hakonensis*, Sai River, 31 July 2012 and 3 June 2013, respectively. (5) Serial sections of livers and hearts (NSMT-PI 5893–5894) of *T. hakonensis*, Sai River, 3 June 2013.

Description (Figs. 1–3). After Shimazu (2007), altered slightly from the present study. Body flat dorsoventrally, elongate, widest at level of anterior part of testis, pointed at anterior extremity, gradually narrowing posteriorly but rounded at posterior extremity, small, 1.29–1.67 by 0.19–0.35 (holotype 1.32 by 0.24) (Fig. 1). Anterior proboscis absent. Tegument spinose; fine setae may be present throughout body; ventrolateral spines present, short transverse rows of one to five spines forming one longitudinal row from near anterior extremity of body to midlevel of cirrus pouch on either side of body, one each in anterior first to sixth transverse rows, thick, 6–8 μm long, two to five (usually four) each in



Figs. 1–4. *Sanguinicola ugui*, adults found in afferent branchial artery of gills and egg found in liver of *Tribolodon hakonensis*. — 1, Holotype (NSMT-PI 5284), entire body, ventral view, excretory system added from free-hand sketches; 2, paratype, anterior part of body, ventral view; 3, holotype, terminal genitalia, dorsal view; 4, fully embryonated egg. Figs. 2–4 redrawn from Shimazu (2007) with slight alteration. Scale bars: 0.5 mm in Fig. 1; 0.1 mm in Figs. 2–3; 0.025 mm in Fig. 4.

remainder, slender, 14–22 μm long (Fig. 2). Nerve chords conspicuous; transverse nerve commissure dorsal to esophagus, 10–12% of body length from anterior extremity. Oral and ventral sucker absent. Mouth small, slightly ventrally subterminal. Small globular sphincter- or sucker-like structure present around mouth,

about 6–9 μm in diameter (Fig. 2). Esophagus narrow, slender, forming fusiform pharynxlike thickening measuring 0.06–0.08 by 0.02–0.04 slightly anteriorly to transverse nerve commissure, surrounded by gland cells anteriorly to ceca (Figs. 1–2). Intestine X-shaped, consisting of usually four but rarely five to six short ceca,

21–23% of body length from anterior extremity. Testis single, elongated longitudinally, median, between ceca and ovary, 0.44–0.60 by 0.09–0.19, with 21–25 lateral lobes on either side. Sperm flowing in dorsal bundle in each lobe, bundles joining together into dorsal median bundle running posteriorly within testis from anterior end of testis to posterior, these bundles apparently lacking any kind of duct. Sperm duct single, running posteriorly from median point of posterior margin of testis to cirrus pouch, ventral to ovary. Cirrus pouch spindle- or club-shaped, thin-walled, 0.14–0.16 by 0.05–0.07, sinistrally submedian, postovarian, directed posteriorly, surrounded by small gland cells, including seminal vesicle and large prostatic cells (Fig. 3). Seminal vesicle club-shaped, thin-walled. Small globular pars prostatica may be differentiated immediately before short eversible ejaculatory duct (cirrus interpreted in Shimazu, 2007). Male genital pore dorsal, sinistrally submedian, some distance from posterior extremity of body, lined with one-layered cuboidal cells, without sphincter around it. Ovary single, bilobed laterally in shape of butterfly, 0.14–0.19 by 0.09–0.14; isthmus 65–71% of body length from anterior extremity. Oviduct originating from posterior margin of ovarian isthmus, passing posteriorly, slightly undulating, dorsal to sperm duct and vitelline duct, including sperm (acting as oviductal seminal receptacle). Seminal receptacle and Laurer's canal absent. Ootype spherical, lined with one-layered columnar cells, 0.03–0.04 in diameter. Large gland cells (Mehlis' gland (?)) seen posterior to ootype, possibly emptying into ootype. Uterus club-shaped, short, 0.06–0.10 by 0.03–0.05, directed posteriorly, bending dorsally at anterior end, lined with one-layered cuboidal cells, surrounded by small gland cells. Sphincter present between ootype and uterus; metraterm not seen. Female genital pore dorsal, dextrally submedian, anterolateral to male genital pore, with well-developed sphincter around it, surrounded by small gland cells. Eggs triangular, one in ootype and 1–7 in uterus if present, 22–34 by 16 μm (collapsed in balsam), not operculate, not embryonated. Vitellaria

follicular, follicles small, profuse mostly between nerve chords from pharynxlike thickening to cirrus pouch, some present laterally to nerve chords, almost confluent anteriorly, separate posteriorly. Common vitelline duct single, beginning at posterior end of pharynxlike thickening, median, in ventral parenchyma; short vitelline ducts joining this duct in places; vitelline reservoir postovarian, in ventral parenchyma, uniting with oviduct to form short ovovitelline duct entering ootype dorsally. Excretory vesicle V-shaped, small, far posterior to ootype; excretory pore posteroterminal.

Excretory system. In living adult specimens, excretory system found arranged asymmetrically (Fig. 1). Right arm of excretory vesicle longer than left. Right main collecting canal long, divided into two short collecting canals between transverse nerve commissure and ceca; left short, divided into two short collecting canals laterally to cirrus pouch. Each collecting canal with one flame cell. Flame cell formula $2 [(1 + 1)] = 4$.

Eggs (Fig. 4). Adults laying unembryonated triangular eggs. Fully embryonated eggs triangular, 40–48 by 34–40 μm in life. In serial sections of liver and heart, eggs found in capillaries (or sinusoids) of lobules of liver, capillaries of cardiac muscle of atrium and ventricle of heart, and capillaries of smooth muscle of arterial bulb of heart, surrounded by granulomatous tissue of host origin of various sizes, usually one egg but rarely two to three eggs per granuloma. No eggs found in gill filaments. Miracidia 30–32 by 18 μm in life, no hatched miracidia seen in either blood vessels or heart.

Remarks. The present study shows that the tegument may be covered by fine cetae throughout the body; the esophagus is surrounded by small gland cells anteriorly to the ceca; and the ovary is slightly larger than described and figured by Shimazu (2007), 0.14–0.19 long (Figs. 1–2) instead of 0.12–0.17 long (figs. 1–2). In addition, Shimazu (2007) misinterpreted the arterial bulb as the ventricle and the afferent branchial artery as the efferent branchial artery. In the figure legend (Shimazu, 2007: 3), “0.2 mm” in Figs. 3 and

5 should be read as "0.1 mm."

Adults were found in the lumen of the blood vessels and heart: the afferent branchial artery of the gills and thick branches of the hepatic artery of the liver, and the arterial bulb of the heart. Shimazu (2007) also recorded worms from the kidneys, but its precise site of infection has not yet been determined. Eggs were triangular (Fig. 4). Embryonated eggs were found in the sinusoids of the lobules of the liver and the capillaries of the cardiac muscle of the ventricle and atrium and smooth muscle of the arterial bulb of the heart, surrounded by a granuloma of host origin. Probably, after being laid in the blood vessels and heart, eggs are carried by blood flow into the capillaries and become fully embryonated there; and granulomas of host origin then develop around them to occlude the capillaries as seen in some other species of *Sanguinicola* (Smith, 1972, 1997). It is interesting that no eggs were found in the gill filaments in the present species in spite of the fact that adult worms lived in the arterial bulb and afferent branchial artery.

Life cycle. Not known. Sanguinicolid cercariae (see Discussion on *Sanguinicola*), which might be identified as *Cercaria andoi* Faust, 1924, were found produced in daughter sporocysts in the snails *Semisulcospira libertina* (Gould, 1859) and *Semisulcospira dolorosa* (Gould, 1859) (Gastropoda, Pleuroceridae) in the Hiroi River. They also had an asymmetrically arranged excretory system as in the adult of *S. ugui* (Shimazu, 2007). Shimazu (2007) failed to infect them to *Tribolodon hakonensis*.

***Sanguinicola hasegawai* sp. nov.**

(Figs. 5–9)

Type host. *Barbatula toni* (Dybowski, 1869) (Nemacheilidae).

Sites of infection. Blood vessels and heart: thick branches of hepatic artery of liver; and atrium, ventricle and arterial bulb of heart.

Type locality. Mamachi River at Aoba (42°49'N, 141°40'E), Chitose City, Hokkaido.

Another locality. Izari River at Eniwa City,

Hokkaido.

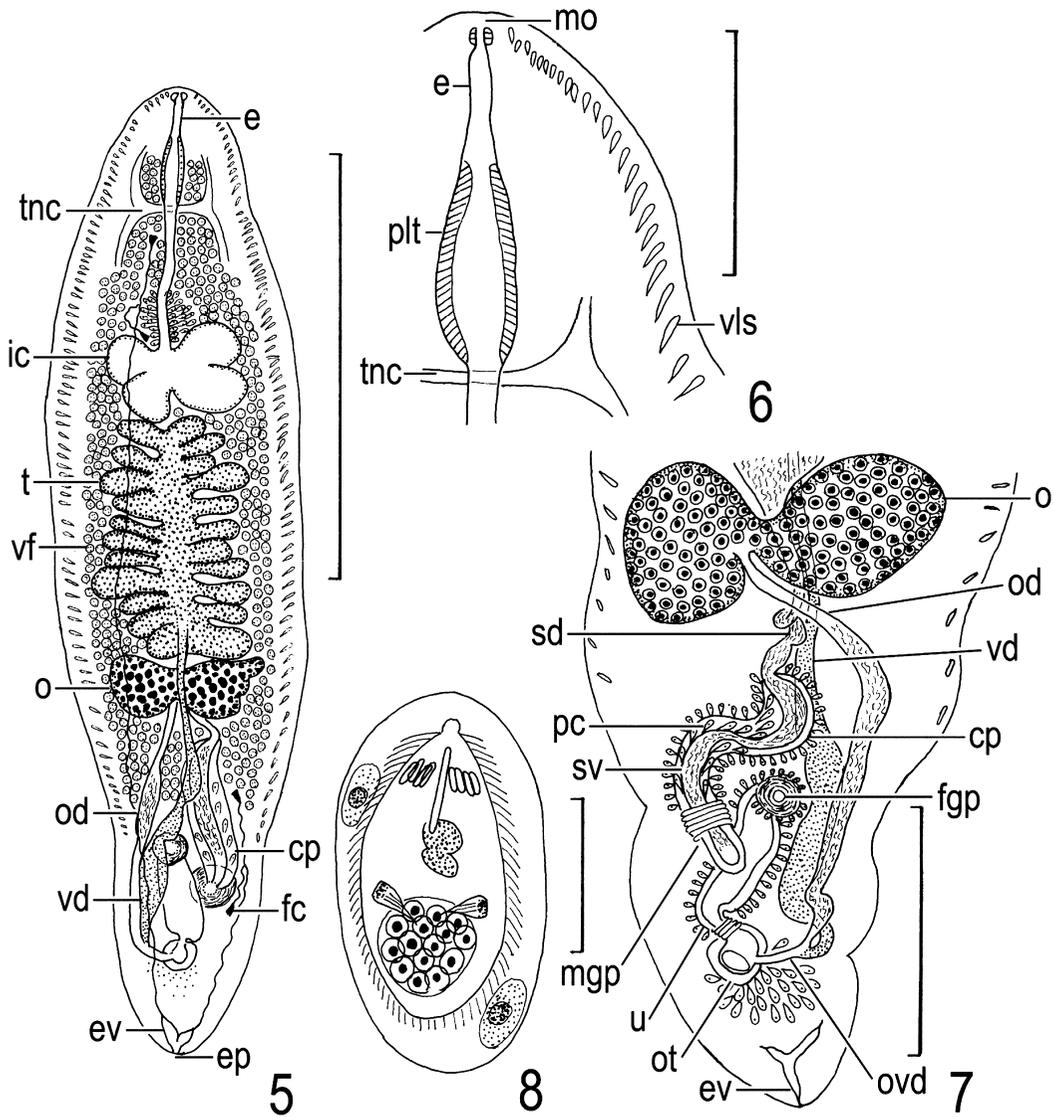
Prevalence and intensity. Not recorded.

Type specimens. (1) Holotype (NSMT-PI 5834), adult, hot formalin-fixed, whole-mounted, ex atrium of *Barbatula toni*, Mamachi River, 20 May 2011. (2) 13 paratypes: 4 (NSMT-PI 5833), adult, flattened, whole-mounted, ex liver of *B. toni*, Mamachi River, 18 April 2011; 2 (NSMT-PI 5835) and 5 (NSMT-PI 5836), adult, hot formalin-fixed, whole-mounted, ex atrium of *B. toni*, Mamachi River, 20 May 2011 and 9 August 2011, respectively; 2 (NSMT-PI 5837), adult, hot formalin-fixed, whole-mounted, ex atrium of *B. toni*, Izari River, 1 June 2011.

Vouchers. 1 specimen (NSMT-PI 5838), adult, hot formalin-fixed, whole-mounted, ex atrium of *B. toni*, Mamachi River, 20 May 2011; 5 (NSMT-PI 5839), adult, hot formalin-fixed, whole-mounted, ex atrium of *B. toni*, Izari River, 1 June 2011; serial sections of livers (NSMT-PI 5840–5841) and hearts (NSMT-PI 5842, 5844) of *B. toni*, Mamachi River, 9 August 2011; and egg masses (NSMT-PI 5843), ex atrium of *B. toni*, Mamachi River, 9 August 2011.

Etymology. The specific name *hasegawai* is dedicated to Dr. Koh Hasegawa, who generously collected the host fish several times for the present study.

Description (Figs. 5–7). Based on the type series; 10 specimens including holotype measured. Similar to the foregoing *Sanguinicola ugui* in general morphology. Body flat dorsoventrally, elongate, widest at level of anterior part of testis, rounded at anterior extremity, gradually narrowing posteriorly but rounded at posterior extremity, small, 0.76–1.13 by 0.19–0.30 (holotype 1.13 by 0.30) (Fig. 5). Fine setae may be present throughout body; ventrolateral spines somewhat curved, lying transversely, large, 8–24 by 2–3 μ m, becoming smaller especially in part of body anterior to transverse nerve commissure, forming single longitudinal row from near anterior extremity to midlevel of cirrus pouch (Figs. 5–6). Transverse nerve commissure 0.12–0.15% of body length from anterior extremity. Sucker-like structure present around mouth, about 8 μ m



Figs. 5–8. *Sanguinicola hasegawai* sp. nov., adults and egg found in *Barbatula toni*. — 5, Holotype (NSMT-PI 5834) found in atrium of heart, entire body, ventral view, excretory system added from free-hand sketches; 6, holotype, anterior part of body, ventral view; 7, paratype (NSMT-PI 5833) found in liver, posterior part of body, dorsal view; 8, fully embryonated egg found in atrium. Scale bars: 0.5 mm in Fig. 5; 0.2 mm in Fig. 7; 0.1 mm in Fig. 6; 0.025 mm in Fig. 8.

in diameter. Pharynxlike thickening of esophagus 0.05–0.08 by 0.02–0.03, immediately anterior to transverse nerve commissure. Small gland cells surrounding esophagus anteriorly to ceca. Ceca four, short, 0.05–0.11 by 0.08–0.17, 29–33% of body length from anterior extremity. Testis 0.20–0.29 by 0.10–0.19, with 9–11 lateral lobes on

either side. Cirrus pouch club-shaped, curved in S-shape, thin-walled, 0.11–0.17 by 0.03–0.05. Seminal vesicle club-shaped, thin-walled. Pars prostatica not clearly seen. Ejaculatory duct may be differentiated, short, eversible (Fig. 7). Male genital pore dorsal, sinistrally submedian, midway in postovarian region of body, encircled by

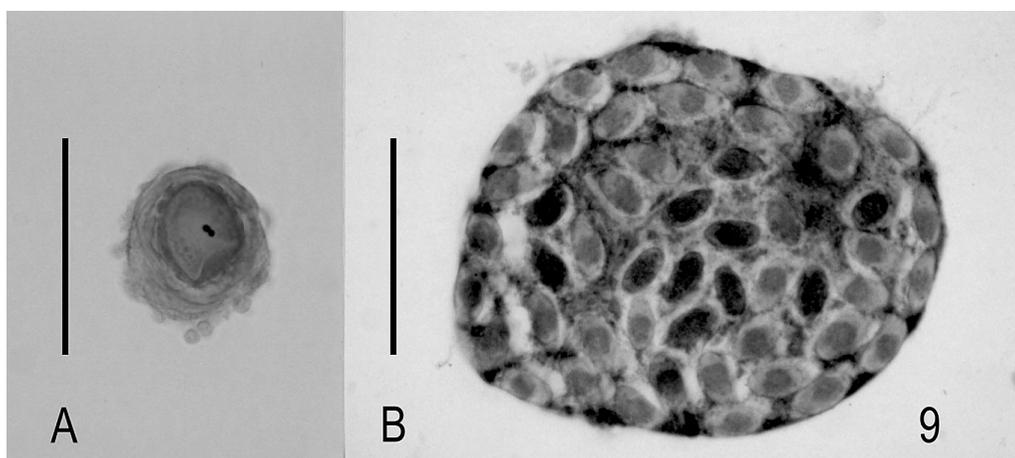


Fig. 9. *Sanguinicola hasegawai* sp. nov. (continued), photomicrographs of eggs (A, solitary egg; B, massed eggs) found in atrium of heart, encapsulated by host cells, stained with Heidenhain's iron hematoxylin. Scale bars: 0.2 mm in B; 0.1 mm in A.

sphincter, surrounded by small gland cells. Ovary bilobed laterally, 0.05–0.08 by 0.09–0.17, lobes globular or slightly irregular in outline; isthmus 62–68% of body length from anterior extremity. Ootype spherical, 0.03–0.05 in diameter, almost median, posterior to male genital pore. Large cells (Mehlis' gland (?)) seen posterior to ootype, possibly emptying into ootype. Uterus clavate, short, 0.05–0.11 by 0.04–0.06. Sphincter present between ootype and uterus. Female genital pore dorsal, dextrally submedian or median, anterior to male genital pore, encircled by well-developed sphincter. Eggs elliptical, thin-shelled, pale-white, not operculate, one in ootype and 1–13 in uterus if present, 27–32 by 13–17 μm , not embryonated. Vitelline follicles profuse mostly between nerve chords from mid-level of pharynxlike thickening to midlevel of cirrus pouch, some present laterally to nerve chords, almost confluent anteriorly, separate posteriorly. Common vitelline duct beginning at midlevel of pharynxlike thickening. Excretory vesicle V-shaped, small, far posterior to ootype; excretory pore posteroterminal.

Excretory system. In living adult specimens, excretory system found arranged asymmetrically (Fig. 5). Right arm of excretory vesicle longer than left. Right main collecting canal long,

divided into two short collecting canals between transverse nerve commissure and ceca; left short, divided into two short collecting canals laterally to cirrus pouch. Each collecting canal with one flame cell. Flame cell formula 2 [(1 + 1)] = 4 or 2 [(2)] = 4

Eggs (Figs. 8–9). Adults laying unembryonated elliptical eggs. Fully embryonated eggs elliptical, 49–60 by 27–36 μm ; miracidia ciliated, 40–48 by 19–25 μm (Fig. 8). Central stylet near anterior extremity of body, 14 by 1 μm ; four small rodlets on either side of it, 5 by 2 μm . Eyespot 8–9 by 5–6 μm , median, slightly pre-equatorial, tightened in middle. Germ cell ball globular, 16–21 μm in diameter, near posterior extremity of body. A pair of flame cells anterior to germ cell ball. No hatched miracidia seen in blood vessels and heart.

After being laid, two or more eggs agglutinated by host cells into masses, both solitary eggs and egg masses encapsulated by host cells in a few layers of various shapes and sizes (Fig. 9A, B) at least in heart and liver, but not in gills: in heart, in lumen of atrium, ventricle and arterial bulb and in lumen of branches of coronary artery; in liver, in lumen of thick branches and arterioles of hepatic artery. No new granulomatous tissue observed around encapsulated eggs

even in arterioles.

Remarks. *Sanguinicola hasegawai* sp. nov. most closely resembles *Sanguinicola megalobrame* Li, 1980 found in *Megalobrama amblycephala* Yih, 1955 (Cyprinidae) from Hubei Province, China (Li, 1980), in having a single longitudinal row of the ventrolateral tegumental spines on either side of the body, an asymmetrically arranged excretory system and elliptical eggs. However, it differs from the latter in having a smaller body, 0.76–1.13 by 0.19–0.30 instead of 1.23–1.74 by 0.23–0.42; no long setae at the posterior part of the body; four short ceca instead of one cecum; fewer lateral lobes of the testis, 9–11 instead of 18–22 on either side; and larger fully embryonated eggs, 49–60 by 27–36 μm instead of 29.5–35.7 by 18.7–20.4 μm .

The asymmetrically arranged excretory system is also known in *Sanguinicola idahoensis* Schell, 1974 found in *Oncorhynchus mykiss* (Walbaum, 1792) (syn. *Salmo gairdneri* Richardson, 1836) (Salmonidae) from Idaho, USA (Schell, 1974); and *S. ugui* found in *Tribolodon hakonensis* (Shimazu, 2007; this paper). *Sanguinicola idahoensis* has elliptical eggs but a larger body, 1.2–2.3 by 0.3–0.5; and more lateral lobes of the testis, 14–18 on either side. *Sanguinicola ugui* has a single longitudinal row of short transverse rows of one to five ventrolateral spines on either side of the body; more lateral lobes of the testis, 21–25 on either side; and triangular eggs.

Adults were found in the lumen of thick branches of the hepatic artery of the liver and the lumen of the atrium, ventricle and arterial bulb of the heart of the host fish. Solitary eggs and egg masses were found encapsulated by host cells in a few layers: in the liver, in the lumen of thick branches and arterioles of the hepatic artery, but not in the lobules; in the heart, in the lumen of the atrium, ventricle and arterial bulb and in the lumen of the branches of the coronary artery. Eggs became fully embryonated in the capsules. No new granulomatous lesions were observed around the encapsulated eggs even in the arterioles. These findings suggest that, after being deposited at the sites of infection of adults, eggs

stand solitarily or two or more eggs become agglutinated into masses by host cells; the solitary eggs and egg masses become encapsulated by host cells; these encapsulated solitary eggs and egg masses are carried by blood flow into the arterioles of the liver and the branches of the coronary artery of the heart to clog them as emboli. It appears that no new granulomas of host origin are added around the emboli. In this histopathology, the present new species differs from some other species of *Sanguinicola*, in which solitary naked eggs are carried by blood flow into the capillaries to lodge there as emboli and then granulomatous tissue of host origin develops around them to occlude the capillaries (see *S. ugui* in this paper). It is not known how fully embryonated eggs or hatched miracidia exit the host fish into the water.

Life cycle. Not known.

***Sanguinicola* sp. of Shimazu, 1999**

Sanguinicolidae gen. sp.: Shimazu, 1999: 67.

Host in Japan. *Acheilognathus tabira tabira* Jordan and Thompson, 1914 (Cyprinidae) (Shimazu, 1999).

Site of infection. Gills (Shimazu, 1999).

Geographical distribution. Shiga Prefecture: Lake Biwa at Onoe, Kohoku-cho, Nagahama City (Shimazu, 1999).

Remarks. An immature specimen of *Sanguinicola* sp. was found in the gill of *A. tabira tabira* on 3 June 1980 (Shimazu, 1999, 2003; Shimazu *et al.*, 2011). This specimen was lost before morphological observations for species identification and description could be undertaken.

Life cycle. Not known.

Discussion on *Sanguinicola*

In *Sanguinicola*, sanguinicolid [or lophocercous-brevifurcate-apharyngeate] cercariae are produced in sporocysts and rediae [(?)] in molluscan intermediate hosts, and cercariae directly penetrate the skin of fish final hosts (Smith,

1972, 1997). Several species of sanguinicolid cercariae have been reported in Japan: *Cercaria andoi* Faust, 1924 and some similar cercariae in *Semisulcospira* spp. (Pleuroceridae); *Cercaria senoi* Faust, 1924 in *Viviparus malleatus* Reeve, 1863 (Viviparidae) and a similar cercaria in *Bullimus manchouricus japonicus* (now *Parafossarulus manchouricus japonicus* (Pilsbry, 1901)) (Stenothyridae); *Cercaria cristhophora* Ito, 1978 in *Austropeplea ollula* (Gould, 1859) and *Lymnaea japonica* (Jay, 1857) (Lymnaeidae) and a similar cercaria in *L. japonica*; and a cercaria in *Sinotaia quadrata historica* (Gould, 1859) (Viviparidae) (Ito, 1964, 1988; Shimazu, 2007). This suggests that, besides *S. ugui* and *S. hasegawai*, some unpublished species of *Sanguinicola* also occur in Japan, though the cercariae themselves need further critical studies.

Judging from my experience, it is not easy to foresee, while examining fish for parasites, which fish harbors small worms of *Sanguinicola* in organs or tissues, especially in case no granulomas caused by laid eggs are seen in the gill filaments as in *S. ugui* and *S. hasegawai*. Fish should be examined as soon as possible after killing, or worms would disappear (perhaps collapse) from the heart and blood vessels. The heart and liver of the fish should be examined first. Worms should be fixed as soon as possible after collecting, or they would disappear (perhaps collapse) in saline in a short time.

The name and authorship of the family, to which *Sanguinicola* belongs, is Aporocotylidae Odhner, 1912 (Bullard *et al.*, 2009), not Sanguinicolidae von Graff, 1907 (Smith, 2002a, b).

Key to two species in *Sanguinicola* in this paper

- 1.1. Ventrolateral tegumental spines arranged in a single longitudinal row on either side of body; testis having 9–11 lateral lobes on either side; eggs elliptical *S. hasegawai* sp. nov.
 1.2. Ventrolateral tegumental spines arranged in a single longitudinal row of short transverse rows of 1–5 spines on either side of body; testis having 21–25 lateral lobes on either side; eggs triangular *S. ugui*

Superfamily **Bivesiculoidea** Yamaguti, 1934

Family **Bivesiculidae** Yamaguti, 1934

***Bivesicula* sp.** of Shimazu, 1994

(Fig. 10)

Bivesicula sp.: Shimazu, 1994: 17, 19, fig. 1.

Host in Japan. *Monopterus albus* (Zuiew, 1793) (Synbranchidae) (Shimazu, 1994).

Site of infection. Intestine (Shimazu, 1994).

Geographical distribution. Okinawa Prefecture: Naha (Shimazu, 1994).

Material examined. 2 specimens (Ozaki's Collection, MPM Coll. No. 30206, labeled "No. *Bivesicula* TAUNAGI Darm [Naha] 1936 No. 13") of *Bivesicula* sp., adult, whole-mounted, ex intestine of *Monopterus albus*, Naha, 1936 (Shimazu, 1994).

Description (Fig. 10). After Shimazu (1994), slightly altered from the present study. Body flattened dorsoventrally, broad-ovate, rounded anteriorly, attenuated posteriorly, small, 1.95–2.00 by 1.25–1.30. Tegumental spines not seen (most likely lost by maceration of body). Eyespot pigment fine, dispersed in forebody. Transverse nerve commissure dorsal to middle of esophagus. Oral and ventral suckers absent. Mouth antero-terminal. Prepharynx not seen. Pharynx globular, 0.12 by 0.14–0.16. Esophagus 0.28–0.31 long, thicker-walled in anterior third (0.08 by 0.02–0.03), surrounded by small gland cells, bifurcating at about junction of first and second fourths of body. Intestinal ceca terminating blindly at about midlevel of testis. Testis single, entire, 0.35–0.43 by 0.43–0.51, median, in third fourth of body. Sperm ducts two, diverging from right

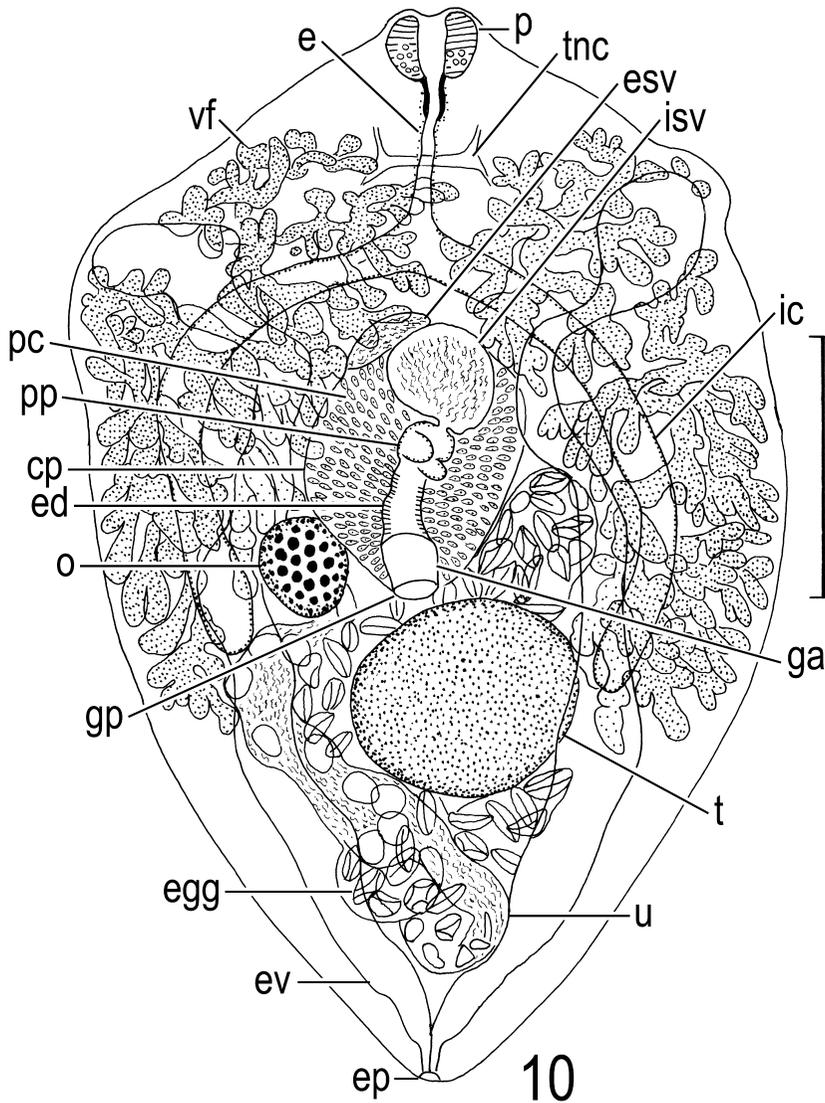


Fig. 10. *Bivesicula* sp., adult found in intestine of *Monopterus albus*, entire body, ventral view, redrawn from Shimazu (1994) with slight alteration. Scale bar: 0.5 mm.

and left anterolateral margin of testis, running dorsally to cirrus pouch. External seminal vesicle oblong, 0.16 by 0.06–0.09, anterodorsal to cirrus pouch. Cirrus pouch elliptical, thin-walled, 0.43–0.50 by 0.35–0.39, median, slightly pre-equatorial, pretesticular. Internal seminal vesicle spherical, constricted posteriorly, 0.16–0.19 by 0.09–0.14. Male terminal duct complex, extending backward. Pars prostatica thick, 0.10–0.12 by 0.09–0.10, with two diverticula, surrounded by

numerous prostatic cells. Ejaculatory duct thick, 0.10 long, surrounded by numerous small gland cells slightly smaller than prostatic cells. Genital atrium may be fairly long. Genital pore on posterior margin of cirrus pouch. Ovary almost globular, 0.16–0.18 by 0.15–0.16, pretesticular, dextrally submedian, posterolateral to cirrus pouch, about level with genital pore. Ovarian complex between ovary and testis, not observed in more detail. Uterus coiled a few times between two

arms of excretory vesicle from near posterior extremity of body to midlevel of cirrus pouch, embracing testis dextrally, acting as uterine seminal receptacle in proximal portion; metraterm not observed clearly. Eggs fairly numerous, thick-shelled, operculate, light brown, 90–100 by 40–60 μm (collapsed), not embryonated. Vitellaria follicular, follicles distributed from midlevel of esophagus to midlevel of testis along ceca, or slightly beyond testis, confluent anteriorly to cirrus pouch, separate posteriorly; most follicles in dorsal and lateral fields of body, some extending ventral to excretory vesicle and ceca. Excretory vesicle V-shaped, large, in ventral parenchyma; arms reaching to midlevel of esophagus; excretory pore posteroterminal.

Remarks. These specimens are somewhat similar to *Bivesicula synodi* Yamaguti, 1938, a marine species, described from *Synodus japonicus* (Houttuyn) [sic, now *Synodus ulae* Schultz, 1953 (Synodontidae) (?)] at Numazu, Shizuoka Prefecture (Yamaguti, 1938); but they are different from the latter in several morphological respects (Shimazu, 1994). They have remained unidentified.

Life cycle. Not known.

Discussion on *Bivesicula*

Species of *Bivesicula* have previously been recorded from marine fishes (Cribb, 2002). *Monopterus albus* is a freshwater fish. It is quite unknown at present whether the present specimens represent a freshwater species of *Bivesicula*. Shimazu (1994) discussed the infection route of the parasite to the host. The freshwater fish *M. albus* devours fishes, worms, crustaceans and other small aquatic animals. If the present specimens belong to a marine species, the occurrence in *M. albus* suggests that *M. albus* acquired infection with this parasite by eating some marine fish or animals that had transferred juvenile or adult worms within them from sea to fresh water. It seems unlikely that planktonic free-living cercariae are directly ingested by *M. albus* after entering fresh water. Otherwise, the

host may have been artificially fed on marine fish harboring worms.

Superfamily **Haploporoidea** Nicoll, 1914

Family **Haploporidae** Nicoll, 1914

Carassotrema koreanum Park, 1938

(Figs. 11–17)

Carassotrema koreanum Park, 1938: 290–292, pl. 13, figs. 1–8; Yamaguti, 1942: 392–393, pl. 24, fig. 4; Shimazu, 2005: 138, figs. 1–3.

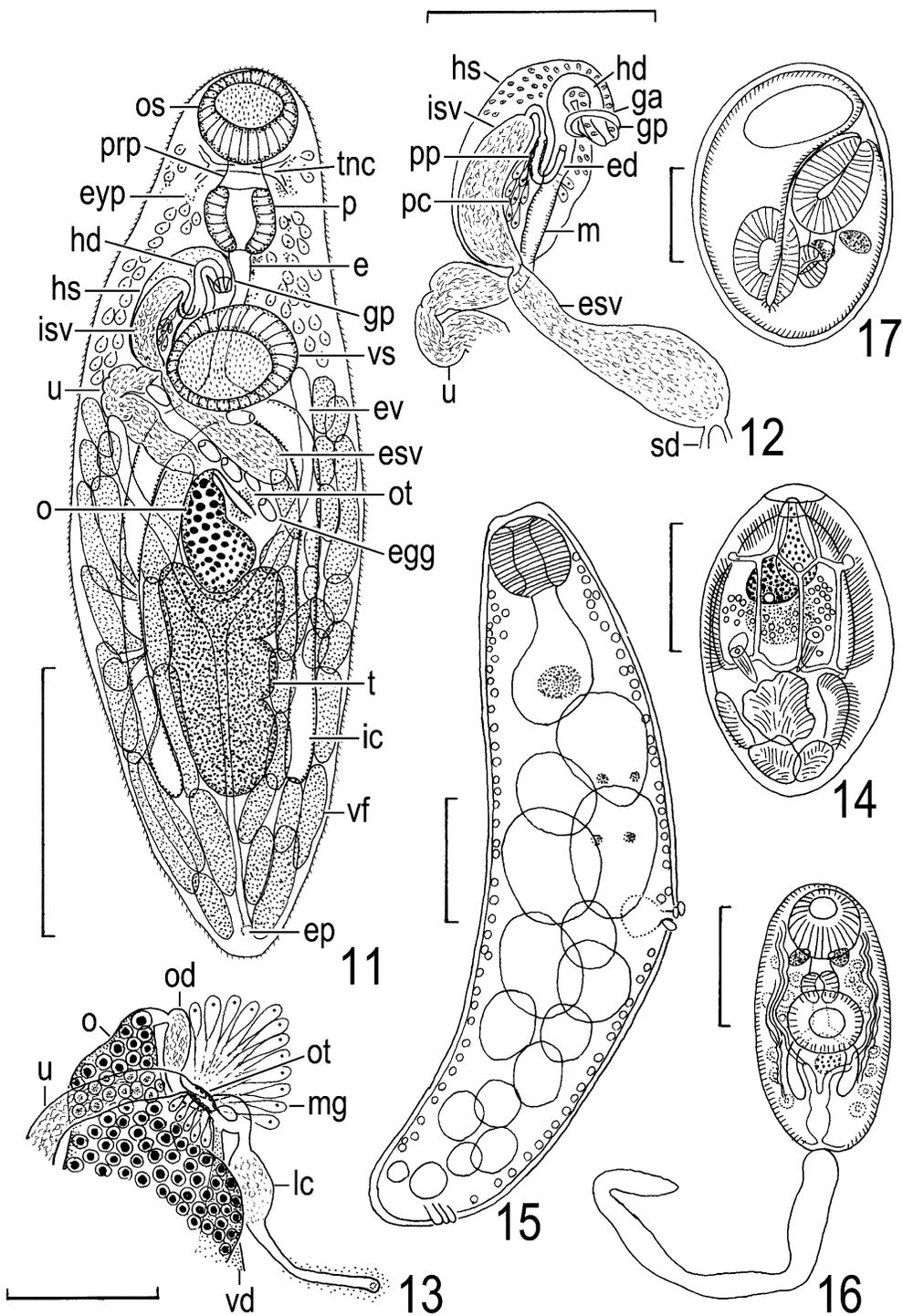
Hosts in Japan. *Cyprinus carpio* Linnaeus, 1758 (Cyprinidae) (Yamaguti, 1942), (?) *Carassius carassius* (Linnaeus, 1758) (Cyprinidae) (Yamaguti, 1942), *Carassius* sp. (Shimazu, 2005; this paper) and *Tribolodon hakonensis* (Cyprinidae) (Shimazu, 2005).

Site of infection. Intestine (Yamaguti, 1942; Shimazu, 2005).

Geographical distribution. (1) Aomori Prefecture: Lake Ogawara at Kamikita Town, now Kamikita-kita, Tohoku Town (Shimazu, 2005). (2) Ibaraki Prefecture: Lake Kasumigaura at Tsuchiura (Yamaguti, 1942; Shimazu, 2005). (3) Osaka Prefecture: Kawachi Province (the southeastern part of the prefecture) (Yamaguti, 1942).

In Primorskiy Kray, Russia (e.g., Bykhovskaya-Pavlovskaya and Kulakova, 1987), Korea (e.g., Park, 1938), China (e.g., Long and Lee, 1958; Tang and Lin, 1979; Wang and Pan, 1984) and Vietnam (?) (e.g., Kulakova and Ha Ky, 1976; Moravec and Sey, 1989).

Material examined. (1) 1 specimen (Yamaguti's Collection, MPM Coll. No. 22270-b, number changed) of *Carassotrema koreanum*, adult, whole-mounted, ex intestine of *Cyprinus carpio*, Lake Kasumigaura, 4 April 1940 (Yamaguti, 1942; Shimazu, 2005). (2) [37] (NSMT-PI 5245) of *C. koreanum*, [4] immature, [33] adult, whole-mounted, ex intestine of *Carassius auratus langsdorfii* (Temminck and Schlegel, 1846) [sic, now *Carassius* sp.], Lake Ogawara, [5–9 September 1997] (Shimazu, 2005). (3) [2] (NSMT-PI 5246) of *C. koreanum*, adult, whole-mounted, ex intestine of *Tribolodon hakonensis*, Lake



Figs. 11–17. *Carassotrema koreanum*, adults found in intestine of *Carassius* sp. and life cycle. — 11, Entire body, ventral view; 12, terminal genitalia, ventral view; 13, ovarian complex, ventral view; 14, fully embryonated egg; 15, [most likely daughter] redia; 16, cercaria; 17, encysted metacercaria. Figs. 14–17 redrawn from Tang and Lin (1979). Scale bars: 0.5 mm in Figs. 11 and 15; 0.3 mm in Fig. 12; 0.1 mm in Figs. 13 and 16; 0.05 mm in Fig. 17; 0.03 mm in Fig. 14.

Ogawara, [4 September 1997] (Shimazu, 2005).

Description (Figs. 11–13). After Shimazu (2005), slightly altered from the present study. Body elongate-oval, small, 1.52–2.26 by 0.60–0.85; forebody 0.51–0.72 long, occupying 32–38% of body length. Tegument spinose, spines dense in anterior part of body, becoming sparser posteriorly. Eyespot pigment scattered on both sides of prepharynx. Numerous large gland cells of two types (with fine and coarse granular cytoplasm, respectively) seen in ventral and dorsal parenchyma, especially in forebody. Transverse nerve commissure dorsal to prepharynx. Oral sucker subglobular, 0.17–0.23 by 0.19–0.24, ventrally subterminal; its rim and inner wall bearing small spines. Prepharynx present, 0.05–0.12 long [small gland cells not seen between oral sucker and prepharynx]. Pharynx globular, large, 0.14–0.19 by 0.14–0.17. Esophagus longer than pharynx, 0.16–0.29 long, surrounded by small gland cells, bifurcating dorsally to ventral sucker. Small gland cells massed together between pharynx and esophagus. Intestinal ceca ending blindly at level of posterior margin of testis or slightly farther, but not reaching to posterior extremity of body. Ventral sucker subglobular, 0.21–0.26 by 0.25–0.30, at about junction of anterior and middle thirds of body; its rim and inner wall bearing small spines; sucker width ratio 1:1.2–1.3. Testis single, cordate, irregularly indented, large, 0.39–0.62 by 0.26–0.55, almost intercecal, located some distance anterior to posterior extremity of body. Sperm ducts two, arising from respective anterolateral corners of testis; common sperm duct absent. External seminal vesicle large, retort- to club-shaped, between ventral sucker and ovary. Hermaphroditic sac elongate, fairly thick-walled, muscular, longer than ventral sucker, 0.23–0.39 by 0.15–0.22, usually dextral or sinistral but rarely dorsal to ventral sucker, usually bent backward in anterior part, containing internal seminal vesicle, prostatic complex, ejaculatory duct and hermaphroditic duct. Internal seminal vesicle elongate, connecting to pars prostatica with short inverted U-shaped male duct. Pars prostatica oblong,

small, surrounded by weakly developed prostatic cells. Ejaculatory duct short, entering lateral wall of metraterm at junction of metraterm and hermaphroditic duct, projecting (forming cirrus of Park, 1938) into hermaphroditic duct. Hermaphroditic duct long, everted, surrounded by small gland cells, apparently being continuation of metraterm in structure. Genital atrium small. Genital pore usually slightly dextral to esophagus between pharynx and ventral sucker. Ovary single, reniform to pyriform, 0.15–0.32 by 0.08–0.16, median or submedian, immediately pretesticular. Ovarian complex anteromedial [not anterolateral] to ovary. Oviduct dilated to contain sperm. Laurer's canal proceeding backward obliquely to open dorsally at ovarian level, dilated in proximal portion to contain sperm (as rudimentary seminal receptacle). Seminal receptacle absent. Ootype small, may not be vesicular; Mehlis' gland large, massive. Uterus coiled a few times on both sides of external seminal vesicle between testis and ventral sucker, overlapping ceca; uterine seminal receptacle well developed; metraterm well developed in hermaphroditic sac. Uterine eggs not numerous, thin-shelled, operculate, light brown, 56–62 by 34–42 μm (collapsed), with small hooklike projection on antiopercular pole, not embryonated. Vitellaria follicular, follicles elliptical to tubular, rarely branched in parts, spreading in lateral fields of hindbody between level of intestinal bifurcation and posterior extremity of body, ventral and dorsal to intestines, separate anteriorly, confluent post-testicularly; vitelline ducts transverse, ventral to ceca, dorsal to testis; common vitelline duct running forward. Excretory vesicle Y-shaped; stem dorsal to testis, bifurcating dorsally to anterior part of testis; arms in ventral parenchyma, extending anteriorly to level of ventral sucker; excretory pore posterodorsal.

Remarks. Park (1938) described *Carassotrema koreanum* as a new genus and species on the basis of adults found in the alimentary canal of *Carassius auratus* (Linnaeus, 1758) caught in the vicinity of Seoul, Korea. Yamaguti (1942) supplemented Park's original description by

describing two and three adult specimens found in the intestine of *Cyprinus carpio* from Lake Kasumigaura and *Ca. carassius* from Kawati [Kawachi] Province (the ancient name of the southeastern part of Osaka Prefecture), respectively, in Japan. Yamaguti had used five specimens then, but Yamaguti's Collection included only one adult specimen (MPM Coll. No. 22270-b) from *Cy. carpio*. Since *Carassius carassius* does not inhabit Japan (Nakabo, 2013), Yamaguti may have examined another species or subspecies of *Carassius* Nilsson, 1832 at that time. Shimazu (2005) described adult specimens found in the intestine of *Carassius* sp. and *Tribolodon hakonensis* from Lake Ogawara.

The present reexamination has shown that the rims and inner walls of the two suckers are armed with small spines (Fig. 11). Park (1938), Yamaguti (1942) and Shimazu (2005) overlooked the spines. Kulakova and Ha Ky (1976) first described that the suckers were spinose in their materials of *C. koreanum* and *C. ginezinskajae* Kulakova and Ha Ky, 1976, not Kha Ki [Ha Ky], 1969, from northern Vietnam. Tang and Lin (1979) presented a figure (fig. 3) to suggest that the two suckers were spinose in their material of *C. koreanum* from China, though they did not mention this. Wang and Pan (1984) illustrated the two smooth suckers in their materials of *C. koreanum* and six other species of *Carassotrema* from China.

Life cycle (Figs. 14–17). Not known in Japan. Tang and Lin (1979) elucidated the life cycle in China. Eggs develop after being laid. Miracidia in fully embryonated eggs have a pigmented eyespot (Fig. 14). The first intermediate host is *Stenothyra toucheana* Heude, 1880 (Gastropoda, Stenothyridae), in which gymnocephalous cercariae (Fig. 16) are produced in [most likely daughter] rediae (Fig. 15). Mother sporocysts were not described. After emerging from the snail host, cercariae encyst upon algae or other tender plants on the bottom of the water. Encysted metacercariae (Fig. 17) were fed to goldfish, from the intestine of which subsequently immature and adult worms of *C. korea-*

num were recovered. Final host fishes are likely to become infected with the parasite by eating algae and tender plants on which metacercariae have encysted. In Japan, Abe in Ueno *et al.* (1930) reported a similar gymnocephalous cercaria (Cercaria D, pl. 1, figs. 7–8) from "*Bithynella*" sp. (Gastropoda, Hydrobiidae) in the neighborhood of Kumamoto [Kumamoto City, Kumamoto Prefecture (?)]. Since Abe stated that this *Bithynella* sp. was very similar to *Parafossarulus manchouricus japonicus* (Pilsbry, 1901) (Stenothyridae) and common in fresh water in Japan, it may have been a stenothyliid.

Discussion on *Carassotrema*

Moravec and Sey (1989) also recorded *Carassotrema koreanum* from northern Vietnam and considered *C. ginezinskajae* to be a junior synonym of *C. koreanum*. However, Moravec and Sey's (1989) material is slightly different from Park's (1938) and the present materials in the egg size, 63–84 by 39–51 μm instead 57–64 by 24–35 μm and 57–72 by 31–38 μm (collapsed), respectively. *Carassotrema ginezinskajae* has eggs measuring 58–68 by 27–43 μm and the vitelline follicles not extending into the extracellular fields of the body (Kulakova and Ha Ky, 1976). These Vietnamese materials need reexamination for definitive species identification.

The oral and ventral suckers are spinose in *C. koreanum* and *C. ginezinskajae* (Kulakova and Ha Ky, 1976; this paper). Tang and Lin (1979) said nothing about this in *C. koreanum* and other five species of *Carassotrema* from China, but their figures (figs. 3 and 17 for *C. koreanum* and fig. 16 for *Carassotrema wui* Tang and Lin, 1963) suggest that the two suckers were spinose. Wang and Pan (1984) illustrated the two smooth suckers in their materials of *C. koreanum* and six other species of *Carassotrema* from China.

A single testis gives off two sperm ducts in *C. koreanum* (Fig. 12, sd). Yamaguti (1942), Kulakova and Ha Ky (1976), Moravec and Sey (1989) and Shimazu (2005) said nothing about the sperm ducts. Park (1938), Long and Lee

(1958) and Tang and Lin (1979) described a single sperm duct. Tang and Lin (1979) also described a single sperm duct in *Carassotrema estuarinum* Tang and Lin, 1979, *C. wui* and *Carassotrema kui* Tang and Lin, 1963 from China. The testis protrudes two anterolateral lobes to various extents. The sperm duct arises out of the anterior corner of each lobe. These suggest that the testicular primordium remains undivided even in the adult stage in *Carassotrema*. The Haploporidae includes a few genera with two testes (Overstreet and Curran, 2005).

Carassotrema wui Tang and Lin, 1963 [sic, should be 1979 (?)], and *Carassotrema kui* Tang and Lin, 1963 [sic, should be 1979 (?)] appear in Tang and Li (1979) and Wang and Pan (1984). Since I have not yet read the abstracts on these two species, I cannot tell at present whether their names are available in accordance with Article 13.1.1 of the International Code of Zoological Nomenclature (ICZN) (International Commission on Zoological Nomenclature, 1999). The known species should be fully redescribed from the type materials, and *Carassotrema* requires a revision (see also Overstreet and Curran, 2005).

Overstreet and Curran (2005) reviewed the Haploporidae. They were wrong in saying eggs as anoperculate and miracidia as lacking eyespots in the diagnosis of *Carassotrema* (see *Life cycle*). They did not refer to the presence of spines on the two suckers. With regards to the author and date of the Haploporidae, they have been cited as "Nicoll, 1914" (e.g., Overstreet and Curran, 2005). Nicoll (1914) used the family name Haploporidae abruptly without discussing it at all. Nicoll's family is identical with the group that Looss (1902) regarded for the time being as a subfamily Haploporinae (Poche, 1926). The author and date of the family and superfamily may be correctly Looss, 1902.

Abbreviations used in the figures. cp, cirrus pouch; e, esophagus; ed, ejaculatory duct; egg, eggs; ep, excretory pore; esv, external seminal vesicle; ev, excretory vesicle; eyp, eyespot pigment; fc, flame cell; fgp, female genital pore; ga,

genital atrium; gp, genital pore; hd, hermaphroditic duct; hs, hermaphroditic sac; ic, intestinal cecum; isv, internal seminal vesicle; lc, Laurer's canal; m, metraterm; mg, Mehlis' gland; mgp, male genital pore; mo, mouth; o, ovary; od, oviduct; os, oral sucker; ot, ootype; ovd, ovovitel-line duct; p, pharynx; pc, prostatic cells; plt, pharynxlike thickening; pp, pars prostatica; prp, prepharynx; sd, sperm duct; sv, seminal vesicle; t, testis; tnc, transverse nerve commissure; u, uterus; vd, vitelline duct; vf, vitelline follicles; vls, ventrolateral spines; vs, ventral sucker.

Acknowledgments

I am grateful to Dr. Koh Hasegawa (Hokkaido National Fisheries Research Institute, Sapporo) and Mr. Satoshi Yamamoto (Nagano Prefectural Fisheries Experimental Station, Azumino) for collecting the host fishes, Dr. Shigehiko Urawa (Hokkaido National Fisheries Research Institute, Sapporo) and Dr. Koji Tojo (Department of Mountain and Environmental Science, Interdisciplinary Graduate School of Science and Technology, Shinshu University, Matsumoto) for laboratory facilities, Dr. Shinpei Wada (Graduate School of Veterinary Medicine and Life Science, Nippon Veterinary and Life Science University, Musashino, Tokyo) for determining the site of infection of *Sanguinicola hasegawai* in the liver of the host, Dr. Toshiaki Kuramochi (NMNS, Tsukuba) and Dr. Takashi Iwaki (MPM, Tokyo) for the loan of the specimens, and Dr. Thomas H. Cribb (School of Biological Sciences, The University of Queensland, Brisbane, Australia) for useful comments on the manuscript.

References

- Abe, H. 1930. [Cercariae of five species and their rediae parasitic in a snail.] *Kumamoto Igakkai Zasshi*, 6: 966–970, pl. 1. (In Japanese.)
- Bray, R. A., D. I. Gibson and A. Jones (eds.) 2008. *Keys to the Trematoda*, 3. xv + 824 pp. CAB International and The Natural History Museum, Wallingford.
- Bullard, S. A., K. Jensen and R. M. Overstreet 2009. Historical account of the two family-group names in use for the single accepted family comprising the "fish

- blood flukes." *Acta Parasitologica*, 54: 78–84.
- Bykhovskaya-Pavlovskaya, I. E. and A. P. Kulakova 1987. [Class TREMATODA — Trematoda Rudolphi, 1808.] In Bauer, O. N. (ed.): [Parasitic Metazoa (Part 2)], 3, pp. 77–198. Izdatel'stvo "Nauka," Leningrad. (In Russian.)
- Cribb, T. H. 2002. Superfamily Bivesiculoidea Yamaguti, 1934. In Gibson, D. I., A. Jones and R. A. Bray (eds.): *Keys to the Trematoda*, 1, pp. 25–29. CAB International and The Natural History Museum, Wallingford.
- Dawes, B. 1956. *The Trematoda with Special Reference to British and Other European Forms*. Reprinted with Corrections. xvi + 644 pp. The Syndics of The Cambridge University Press, London.
- Eschmeyer, W. N. (ed.) 2013. *Catalog of Fishes*. California Academy of Sciences (<http://research.calacademy.org/research/ichthyology/catalog/fishcatmain.asp>). Electronic version accessed 8 August 2013.
- Gibson, D. I., A. Jones and R. A. Bray (eds.) 2002. *Keys to the Trematoda*, 1. xiv + 521 pp. CAB International and The Natural History Museum, Wallingford.
- Goto, A. 1987. [Freshwater fishes — grouping based on their life cycles and formation of their distribution areas.] In Mizuno, N. and A. Goto: [Freshwater Fishes of Japan], pp. 1–15. Tokai University Press, Tokyo. (In Japanese.)
- International Commission on Zoological Nomenclature 1999. *International Code of Zoological Nomenclature*, Fourth Edition. 306 pp. International Trust for Zoological Nomenclature, London.
- Ito, J. 1964. A monograph of cercariae in Japan and adjacent territories. In Morishita, K., Y. Komiya and H. Matsubayashi (eds.): *Progress of Medical Parasitology in Japan*, 1, pp. 389–550. Meguro Parasitological Museum, Tokyo.
- Ito, J. 1988. A subsequent monograph of cercariae in Japan (1962–1988). *Japanese Journal of Parasitology*, 37: 269–322.
- Jones, A. 2005. Superfamily Haploporoidea Nicoll, 1914. In Jones, A., R. A. Bray and D. I. Gibson (eds.): *Keys to the Trematoda*, 2, pp. 127–128. CAB International and The Natural History Museum, Wallingford.
- Jones, A., R. A. Bray and D. I. Gibson (eds.) 2005. *Keys to the Trematoda*, 2. xvi + 745 pp. CAB International and The Natural History Museum, Wallingford.
- Kamegai, S. and A. Ichihara 1972. A check list of the helminths from Japan and adjacent areas Part I. Fish parasites reported by S. Yamaguti from Japanese waters and adjacent areas. *Research Bulletin of the Meguro Parasitological Museum*, (6): 1–40.
- Kha Ki [Ha Ky] 1969. [Parasite fauna of some freshwater fishes in North Vietnam and measures against the most important fish diseases.] 18 pp. Avtoreferat Dissertatsii na Soiskanie Uchevoy Stepeni Kandidata Biologicheskikh Nauk. Akademiya Nauk SSSR, Zoologicheskii Institut, Leningrad. (In Russian.)
- Komiya, Y. 1965. Metacercariae in Japan and adjacent territories. In Morishita, K., Y. Komiya and H. Matsubayashi (eds.): *Progress of Medical Parasitology in Japan*, 2, 328 pp. Meguro Parasitological Museum, Tokyo.
- Kulakova, A. P. and Ha Ky 1976. *Carassotrema koreanum* Park, 1938 (Trematoda, Waretrematidae) and a new species of this genus from freshwater fishes of North Viet-Nam. *Parazitologiya*, 10: 460–462. (In Russian with English abstract.)
- Li, L.-X. 1980. Studies on the agency of sanguinicosis of blunt-snout bream (*Megalobrama amblycephala*) and its control, with description of a new species. *Journal of Fisheries of China*, 4: 179–196, pls. 1–4. (In Chinese with English abstract.)
- Long, S. and W.-C. Lee 1958. Parasitic worms from Tai Hu fishes: digenetic trematodes. II. Opisthorchiidae and other families, with a description of a new species of *Opisthorchis*. *Acta Zoologica Sinica*, 10: 369–376. (In Chinese with English abstract.)
- Looss, A. 1902. Die Distomen — Unterfamilie der *Haploporinae*. *Archives de Parasitologie*, 6: 129–143.
- Moravec, F. and O. Sey 1989. Some trematodes of freshwater fishes from North Vietnam with a list of recorded endohelminths by fish hosts. *Folia Parasitologica*, 36: 243–262.
- Nakabo, T. (ed.) 2013. *Fishes of Japan with Pictorial Keys to the Species*, Third Edition. xlix + xxxii + xvi + 2428 pp. Tokai University Press, Hadano. (In Japanese.)
- Nicoll, W. 1914. The trematode parasites of fishes from the English Channel. *Journal of the Marine Biological Association of the United Kingdom*, 10: 466–505.
- Overstreet, R. M. and S. S. Curran 2005. Family Haploporidae Nicoll, 1914. In Jones, A., R. A. Bray and D. I. Gibson (eds.): *Keys to the Trematoda*, 2, pp. 129–165. CAB International and The Natural History Museum, Wallingford.
- Park, J. T. 1938. A new fish trematode with single testis from Korea. *Keijo Journal of Medicine*, 9: 290–298, pl. 13.
- Poche, F. 1925 [issued 1926]. Das System der Platyhelminthes. *Archiv für Naturgeschichte, Abteilung A, Zoology*, 91: 1–458, fig., plates 1–7.
- Schell, S. C. 1974. The life history of *Sanguinicola idahoensis* sp. n. (Trematoda: Sanguinicolidae), a blood parasite of steelhead trout, *Salmo gairdneri* Richardson. *Journal of Parasitology*, 60: 561–566.
- Shimazu, T. 1994. A species of *Bivesicula* (Digenea: Bivesiculidae) from a freshwater fish of Japan. *Journal of Nagano Prefectural College*, (49): 17–20.
- Shimazu, T. 1995. A revised checklist and bibliography of the platyhelminth parasites reported by Dr. Yoshimasa Ozaki, 1923–1966, and their specimens deposited in the Meguro Parasitological Museum, Tokyo. *Journal of*

- Nagano Prefectural College, (50): 33–50.
- Shimazu, T. 1999. [Turbellarians and trematodes of freshwater animals in Japan.] In Otsuru, M., S. Kamegai and S. Hayashi (eds.): *Progress of Medical Parasitology in Japan*, 6, pp. 65–86. Meguro Parasitological Museum, Tokyo. (In Japanese.)
- Shimazu, T. 2003. Turbellarians and trematodes of freshwater animals in Japan. In Otsuru, M., S. Kamegai and S. Hayashi (eds.): *Progress of Medical Parasitology in Japan*, 7, pp. 63–86. Meguro Parasitological Museum, Tokyo.
- Shimazu, T. 2005. Digeneans found in fresh- and brackish-water fishes of Lake Ogawara in Aomori Prefecture, Japan. *Bulletin of the National Science Museum, Tokyo, Series A (Zoology)*, 31: 137–150.
- Shimazu, T. 2007. Digeneans (Trematoda) of freshwater fishes from Nagano Prefecture, central Japan. *Bulletin of the National Museum of Nature and Science, Series A (Zoology)*, 33: 1–30.
- Shimazu, T. and J. Araki 2006. A list of the helminth parasite specimens deposited in the Department of Zoology, the University of Museum, the University of Tokyo. *The University Museum, The University of Tokyo, Material Reports*, (62): 151–161.
- Shimazu, T., M. Urabe and M. J. Grygier 2011. Digeneans (Trematoda) parasitic in freshwater fishes (Osteichthyes) of the Lake Biwa basin in Shiga Prefecture, central Honshu, Japan. *National Museum of Nature and Science Monographs*, (43): 1–105.
- Smith, J. W. 1972. The blood flukes (Digenea: Sanguinicolidae and Spirorchidae) of cold-blooded vertebrates and some comparison with the schistosomes. *Helminthological Abstracts, Series A*, 41: 161–204.
- Smith, J. W. 1997. The blood flukes (Digenea: Sanguinicolidae and Spirorchidae) of cold-blooded vertebrates: Part 1. A review of the literature published since 1971, and bibliography. *Helminthological Abstracts*, 66: 255–294.
- Smith, J. W. 2002a. Superfamily Schistosomatoidea Stiles and Hassall, 1898. In Gibson, D. I., A. Jones and R. A. Bray (eds.): *Keys to the Trematoda*, 1, pp. 415–417. CAB International and The Natural History Museum, Wallingford.
- Smith, J. W. 2002b. Family Sanguinicolidae von Graff, 1907. In Gibson, D. I., A. Jones and R. A. Bray (eds.): *Keys to the Trematoda*, 1, 433–452. CAB International and The Natural History Museum, Wallingford.
- Tang, Z.-Z. and H.-M. Lin 1979. Studies on *Carassotrema* Park, 1938. Life-histories and distribution. *Xiamen Daxue Xuebao*, 1979: 81–98. (In Chinese with English abstract.)
- Wang, W.-J. and J.-p. Pan 1984. Studies on the *Carassotrema* Park, 1938 (Digenea: Haploporidae Nicoll, 1914) in China. In Institute of Hydrobiology Academia Sinica (ed.): *Parasitic Organisms of Freshwater Fish of China*, pp. 139–148. Agricultural Publishing House, Beijing. (In Chinese.)
- Yamaguti, S. 1938. Studies on the Helminth Fauna of Japan. Part 21. Trematodes of Fishes, IV. 139 pp. + 1 pl. Published by author, Kyoto.
- Yamaguti, S. 1942. Studies on the helminth fauna of Japan. Part 39. Trematodes of fishes mainly from Naha. *Transactions of the Biogeographical Society of Japan*, 3: 329–398, pl. 24.