# Reconstruction of Flower Color of *Amherstia nobilis* by *in vitro* Examination

# Takayuki Mizuno<sup>1</sup>, Nobuyuki Tanaka<sup>1</sup>, Mu Mu Aung<sup>2</sup>, Tomohisa Yukawa<sup>1</sup> and Tsukasa Iwashina<sup>1,\*</sup>

 <sup>1</sup>Department of Botany, National Museum of Nature and Science, 4–1–1 Amakubo, Tsukuba, Ibaraki 305–0005, Japan
<sup>2</sup> Forest Research Institute, Forest Department, Ministry of Natural Resources and Environmental Conservation, Yezin, Nay Pyi Taw, Myanmar
\*E-mail: iwashina@kahaku.go.jp

(Received 17 July 2019; accepted 25 September 2019)

**Abstract** Flower anthocyanin of *Amherstia nobilis* has been reported as pelargonidin 3-*O*-pentoside. However, they were more recently identified as cyanidin 3-*O*-glucoside and 3-*O*-xyloside, and peonidin 3-*O*-glucoside, together with eight flavonol glycosides, i.e. isorhamnetin 3-*O*-glucoside, 7-*O*-glucoside, 3,7-di-*O*-glucoside and 3-*O*-rutinoside, quercetin 3-*O*-rutinoside and 3-*O*-glucocoside, and kaempferol 3-*O*-rutinoside and 3-*O*-glucoside. Color of cyanidin 3-*O*-glucoside which is major anthocyanin in the flowers is originally red but not reddish orange. We presumed that color of anthocyanins is hypsochromically shifted under lower pH, and measured pH of the pressed juice of the petals. As the results, it was proved that pH of the petals is lower (pH 3.3) and in spite of flower pigments are cyanidin glycosides, the flowers of *A. nobilis* express reddish orange. This was followed by *in vitro* examination.

Keywords: *Amherstia nobilis*, cell sap pH, cyanidin 3-*O*-glucoside, *in vitro* examination, reddish orange flower color.

#### Introduction

The genus *Amherstia* (Leguminosae) consists of only one species, *A. nobilis* Wall. and is endemic to Myanmar. Although the species is growing in some botanical gardens in tropical area and greenhouses, its wild population in Myanmar is not known. *A. nobilis* is generally regarded as one of the most beautiful of all flowering trees (Fig. 1) (Bisby *et al.*, 1994). Its flower anthocyanin has been reported to be pelargonidin 3-*O*-pentoside without chemical data (Beale *et al.*, 1941). More recently, three anthocyanins and eight flavonols were isolated from the reddish orange flowers of this species. However, anthocyanins were identified as cyanidin 3-*O*-glucoside and 3-O-xyloside, and peonidin 3-O-glucoside (Iwashina *et al.*, 2019). On the other hand, flavonols were characterized as isorhamnetin 3-O-glucoside, 7-O-glucoside, 3,7-di-O-glucoside and 3-O-rutinoside, quercetin 3-O-rutinoside and 3-O-glucoside, and kaempferol 3-O-rutinoside and 3-O-glucoside (Iwashina *et al.*, 2019). Although major anthocyanin was cyanidin 3-O-glucoside, its flower color was reddish orange which is such as due to pelargonidin and/ or carotenoids. In this survey, we clarified the coloring mechanism of reddish orange flowers by pH measurement of the petals and *in vitro* examination.

<sup>© 2019</sup> National Museum of Nature and Science



Fig. 1. Flowers of *Amherstia nobilis* (photographed in Bogor Botanic Garden, Indonesia by one of the author, T. Iwashina in 1 July 2016).

# **Materials and Methods**

## Plant materials

The flowers were collected in the greenhouse of the Tsukuba Botanical Garden, National Museum of Nature and Science, Tsukuba, Japan in 21 January 2016.

#### Isolation and identification

Flowers were extracted with MeOH/HCOOH (92:8). Anthocyanins and flavonols in crude extracts were characterized by HPLC comparisons with authentic samples which were obtained from the flowers of this species (Iwashina *et al.*, 2019). HPLC measurement was performed using Shimadzu HPLC system (Shimadzu Co., Ltd., Kyoto,), Inertsil ODS-4 column (I.D. 6.0  $\times$  150 mm, GL Sciences Inc., Tokyo), at flow-rate of 1.0 ml min<sup>-1</sup>, detection wave length of 530 nm for anthocyanins and 350 nm for flavonols, and eluents were MeCN/HOAc/H<sub>2</sub>O/H<sub>3</sub>PO<sub>4</sub> (10:8:79:3) for anthocyanins and MeCN/H<sub>2</sub>O/H<sub>3</sub>PO<sub>4</sub> (20:80:0.2) for flavonols.

#### pH measurement

The pH values of the pressed juice from the flower tissues were measured using a handhold pH meter (Twin pH; AS-212, Horiba Instruments, Kyoto). The reddish orange epidermis was carefully peeled and placed between glasses, and pressed to get the pressed juice. The pH of the pressed juice prepared was immediately measured. Measurement were triplicated.

## In vitro examination

The pH effect of cyanidin 3-O-glucoside (5  $\times$  10<sup>-4</sup> M) was determined at different pH values (pH 2.6, 3.3, 4.3, 5.3 and 6.3) of Mellvaine buffer prepared by mixing 0.1 M citric acid and 0.2 M Na<sub>2</sub>HPO<sub>4</sub>. The absorption spectra of the solution were measured immediately (0min) and after 30 min.

## **Results and Discussion**

Flower color of *Amherstia nobilis* was reddish orange, presumed that it is due to carotenoid and/

or anthocyanin, especially pelargonidin. Practically, pelargonidin 3-O-pentoside has been reported as flower pigment without chemical data (Beale et al., 1941). However, cyanidin 3-O-glucoside was found in the flowers of this species as a major pigment, together with minor cyanidin 3-O-xyloside and peonidin 3-O-glucoside, and eight flavonol glycosides, instead of pelargonidin glycoside and carotenoids (Iwashina et al., 2019). Although cvanidin glycosides commonly express red color in almost flowers except for a few plants, e.g. Meconopsis spp. (Takeda et al., 1996; Tanaka et al., 2001; Yoshida et al., 2006) and Centaurea cyanus L. (Takeda et al., 2005; Shiono et al., 2005), reddish orange color was expressed by cyanidin 3-O-glucoside in A. nobilis flowers. Such the fact was presumed that other factors are changed the pigment color (red) to reddish orange in this flowers. Generally, because color of anthocyanins is hypsochromically shifted under lower pH (acidity), pH of the pressed juice of the flowers was measured. As the results, it was shown that pH of the petals is lower (pH 3.3) and in spite of major flower pigment is cyanidin 3-O-glucoside, the flowers of A. nobilis expressed reddish orange. It was followed by in vitro examination (Table 1 and Fig. 2). Absorption maxima of intact petals was  $\lambda$  max 511 nm (Fig. 2A). In in vitro examination, absorption maxima of cyanidin 3-O-glucoside were measured in conditions of pH 2.6, 3.3, 4.3,

5.3 and 6.3 (Table 1 and Fig. 2). In lower acidic solutions (pH 2.6 and 3.3), their absorption maxima showed  $\lambda$ max 511 and 512 nm without additional absorption peaks and shoulders, which was the same with that of intact petals. On the other hand, absorption maxima,  $\lambda max 519 \, nm$  was shown in pH 4.3. Moreover, absorption maxima appeared as two peaks (434 and 529 nm, and 439 and 535 nm, respectively) in pH 5.3 and 6.3 (Fig. 2B). Although absorption maxima were measured after 30 min, those of pH 2.6 and 3.3 were very stable. However, those of pH 5.3 and 6.3 were unstable (Fig. 2C). Thus, it was proved that reddish orange color of A. nobilis flowers is due to lower acidic condition in cell sap. Accompanyingly, although absorption maxima of cyanidin 3-O-glucoside were measured in addition to isorhamnetin 3-O-glucoside which was one of the major flavonols in the flowers of A. nobilis, the bathochromic and also hypsochromic shifts were

Table 1. Absorption maxima of cyanidin 3-Oglucoside with various pH

рН	$\lambda \max(nm)$	
	0 min	30 min
2.6	511	511
3.3	512	511
4.3	519	510
5.3	434, 529	514
6.3	439, 535	438, 533

 $\lambda$ max of fresh petal of *Amherstia nobilis* was shown at 511 nm.

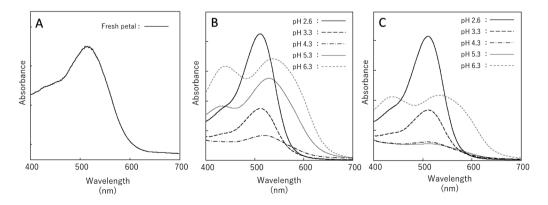


Fig. 2. Absorption spectra of intact petal (A), cyanidin 3-*O*-glucoside with various pH (0 min, B, and after 30 min C).

hardly occurred (data not shown).

The pH of flower cell sap is generally slightly acidic (Shibata et al., 1949). However, pH of the pigmented epidermal cell sap of A. nobilis was found to be extremely lower. The pH of epidermis of the blue and greenish blue petals of morning glory cv. Heavenly Blue and jade vain (Strongylodon macrobotrys A. Gray) were reported to be slightly alkali (pH 7.5) (Stewart et al., 1975) and pH 7.9 (Takeda et al., 2010), respectively. By contraries, pH of the flower cells of Begonia cv. Orange Schwabenland and Azalea cv. Gloria were lower pH (2.5 and 3.1, respectively). Their flower colors were strong redorange and light yellowish pink in spite of their anthocyanins are cyanidin glycosides (Stewart et al., 1975). Although major pigment of A. nobilis was cyanidin 3-O-glucoside, its flower color is changed to reddish orange due to lower acidity of cell saps. There is close correlation between flower color and pollinators (Harborne and Grayer, 1994). Flavonol glycosides directly do not influence the flower color of A. nobilis. However, since insects and birds can discriminate UV range, flavonol glycosides may be useful as pollinator attractants. Although wild population of A. nobilis in Myanmar is unknown, we presume that unknown insects, birds or animals, which were tempted to reddish orange color, visit to this flowers.

#### References

- Beale, G. H., Price, J. R. and Sturgess, V. C. 1941. A survey of anthocyanins VII. The natural selection of flower colour. Proceedings of the Royal Society of London, Series B 130: 113–126.
- Bisby, F. A., Buckingham, J. and Harborne, J. B. (Eds.)

1994. Phytochemical Dictionary of the Leguminosae. Volume 1. Plants and their constituents. pp. 66. Chapman & Hall, London.

- Harborne, J. B. and Grayer, R. J. 1994. Flavonoids and insects. In: Harborne, J. B. (Ed.), The Flavonoids. Advances in Research since 1986. pp. 589–618. Chapman & Hall, London.
- Iwashina, T., Tanaka, N., Aung, M. M., Mizuno, T. and Yukawa, T. 2019. Anthocyanins and flavonols from the flowers of *Amherstia nobilis* endemic to Myanmar. Biochemical Systematics and Ecology 86: https://doi. org/10.1016/j.bse.2019.05.014.
- Shibata, K., Hayashi, K. and Isaka, T. 1949. Über Wasserstoffionenkonzentration des Pressaftes von den anthocyan-führenden Pflanzenorganen. Acta Phytochimica 15: 17–33.
- Shiono, M., Matsugaki, N. and Takeda, K. 2005. Structure of the blue cornflower pigment. Nature 436: 791.
- Stewart, R. N., Norris, K. H. and Asen, S. 1975. Microspectrophotomeric measurement of pH and pH effect on color of petal epidermal cells. Phytochemistry 14: 937–942.
- Takeda, K., Fujii, A., Senda, Y. and Iwashina, T. 2010. Greenish blue flower colour of *Strongylodon macrobotrys*. Biochemical Systematics and Ecology 38: 630– 633.
- Takeda, K., Yamaguchi, S., Iwata, K., Ysujino, Y., Fujimori, T. and Fusain, S. Z. 1996. A malonylated anthocyanin and flavonols in the blue flowers of *Meconopsis*. Phytochemistry 42: 863–865.
- Takeda, K., Osakabe, A., Saito, S., Furuyama, D., Tomita, A., Kojima, Y., Yamadera, M. and Sakuta, M. 2005. Components of protocyanin, a blue pigment from the blue flowers of *Centaurea cyanus*. Phytochemistry 66: 1607–1613.
- Tanaka, M., Fujimori, T., Uchida, I., Yamaguchi, S. and Takeda, K. 2001. A malonylated anthocyanin and flavonols in blue *Meconopsis* flowers. Phytochemistry 56: 373–376.
- Yoshida, K., Kitahara, S., Ito, D. and Kondo, T. 2006. Ferric ions involved in the flower color development of the Himalayan blue poppy, *Meconopsis grandis*. Phytochemistry 67: 992–998.